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# Sensitivity and flexibility of regular and chaotic calcium oscillations

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## Abstract

Sensitivity and flexibility are important properties of biological systems. These properties are here investigated for intracellular calcium oscillations. For a particular model, we comparatively investigate sensitivity and flexibility of regular and chaotic  $\text{Ca}^{2+}$  oscillations. For this model, we obtain two main results. First, sensitivity of the model system to parameter shifting does not depend on the complexity of  $\text{Ca}^{2+}$  oscillations. We observe, however, that both regular and chaotic  $\text{Ca}^{2+}$  oscillations are highly sensitive in regions close to bifurcation points. Second, also flexibility of  $\text{Ca}^{2+}$  oscillations does not significantly depend on the type of  $\text{Ca}^{2+}$  oscillations. Our results show that regular as well as chaotic  $\text{Ca}^{2+}$  oscillations in the studied model are highly flexible in regimes with weak dissipation. Both results are discussed in the sense of possible biological importance.

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## 1. Introduction

The importance of cytosolic calcium is well established in a large variety of cell types. In excitable as well as in non-excitable cells, a significant part of signal transduction from receptors at the cell membrane to enzymes, controlling the complex behaviour of the biological systems, is performed by the oscillatory changing in free cytosolic  $\text{Ca}^{2+}$  concentration, the so-called  $\text{Ca}^{2+}$  oscillations. They regulate many cellular processes from egg fertilisation to cell death [1]. The mechanisms of these oscillations have been intensely

investigated both from experimental and theoretical point of view (for review see [2,3]).

Calcium has to play a multiplicity of roles in order to trigger different cellular functions [1]. Therefore, flexible, yet precisely regulated, information encoding of  $\text{Ca}^{2+}$  oscillations in their frequency [4–10] as well as in their amplitude [11,12] is required. Thus for reliable functioning, a biological system has to be stable, highly sensitive and flexible. Suguna et al. [13] showed that a minimal condition for a model system to be stable, highly sensitive and flexible is a combination of a pair of coupled negative and positive feedback processes. However, the question remains under what conditions, determined by the model parameters, the system is most sensitive and flexible.

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Some authors have already dealt with similar questions. Kummer et al. [14], L  er et al. [15] and Zhong et al. [16] have investigated the impact of noise on the sensitivity of  $\text{Ca}^{2+}$  oscillations. It has been shown that adding Gaussian noise to a subthreshold extracellular stimulus causes noise-induced  $\text{Ca}^{2+}$  oscillations. This means that noise increases the sensitivity in the dose-response relation of calcium, which may have important physiological consequences in enhancing the detection of weak input signals. Galvanovskis and Sandblom [17,18] also studied the sensitivity of  $\text{Ca}^{2+}$  oscillations in response to weak external forcing and noise. Their experimental results showed influences of low frequency electromagnetic fields on the behaviour of living systems. They also studied this phenomenon theoretically on two mathematical models of intracellular  $\text{Ca}^{2+}$  oscillations: the two-variable model of Goldbeter et al. [19] and the three-variable model proposed by Shen and Larter [20]. In the latter, the sensitivity of regular and chaotic regimes was compared. Results indicated higher sensitivity of chaotic  $\text{Ca}^{2+}$  oscillations to variation of parameter values, suggesting a possible role of chaotic processes in detection of weak signals within cells.

The aim of the present study is to analyse the interrelation between sensitivity and flexibility of the same model system as studied by Sandblom and Galvanovskis (i.e. the model proposed by Shen and Larter [20]) and to determine the conditions under which the model is extremely sensitive and flexible. In particular, sensitivity and flexibility of regular and chaotic regimes are compared. Sensitivity is quantified by changes in the model variables caused by shifting the parameter values. We take use of the so-called response coefficients, which were defined in the theoretical framework of the metabolic control analysis (see e.g. [21]) and afterwards successfully applied to the control analysis of different periodic phenomena in biological systems (see e.g. [22,23]). For better comparison of our results to that obtained earlier by Galvanovskis and Sandblom [17] for the same model system, we use a slightly modified definition of the response coefficients. We shift the parameter values of the model system and calculate the ratio of the power of modulated

$\text{Ca}^{2+}$  oscillations over the power of non-modulated  $\text{Ca}^{2+}$  oscillations. This gives information about the energy change of the signal related mainly to its amplitude, however, not so much to its frequency. Therefore, we additionally examine the sensitivity of the system regarding changes in the oscillation frequency, by shifting the parameter values and calculating the ratio of the dominant oscillation frequency of modulated  $\text{Ca}^{2+}$  oscillations over the dominant oscillation frequency of non-modulated  $\text{Ca}^{2+}$  oscillations. Flexibility of the model system is studied by applying external periodic forcing to the model and analysing the changes in frequency of  $\text{Ca}^{2+}$  oscillations. The span of the frequency range in which  $\text{Ca}^{2+}$  oscillations are synchronised with the forcing signal represents the system ability to adapt the basic  $\text{Ca}^{2+}$  oscillations to the external forcing signal. Thus, the span of the frequency range is taken as a measure for the flexibility of the system.

## 2. Mathematical model

We analyse sensitivity and flexibility of the mathematical model proposed by Shen and Larter [20]. The functioning of the model system is based on the mechanisms of calcium-induced calcium release (CICR) [24] and the inositol trisphosphate crosscoupling (ICC) [25,26]. The ICC and CICR mechanisms provide two positive feedbacks (for details see [20]). There are three variables in the model: free  $\text{Ca}^{2+}$  concentration in the cytosol ( $\text{Ca}_{\text{cyt}}$ ), free  $\text{Ca}^{2+}$  concentration in the ER ( $\text{Ca}_{\text{er}}$ ), and the inositol trisphosphate concentration in the cytosol ( $\text{IP}_3$ ). The evolution of the model system is governed by the following differential equations (for parameter values see Table 1):

$$\frac{d\text{Ca}_{\text{cyt}}}{dt} = J_{\text{ch}} + J_{\text{leak}} - J_{\text{pump}} + J_{\text{in}} - J_{\text{out}}, \quad (1)$$

$$\frac{d\text{Ca}_{\text{er}}}{dt} = J_{\text{pump}} - J_{\text{ch}} - J_{\text{leak}}, \quad (2)$$

$$\frac{d\text{IP}_3}{dt} = J_{+} - J_{-}, \quad (3)$$

Table 1  
Model parameters for which all results are calculated unless otherwise stated

Parameter	Meaning	Value
$k_{\text{ch}}$	Maximal rate constant of $\text{Ca}^{2+}$ channels in the ER membrane	$3000.0 \mu\text{M s}^{-1}$
$k_{\text{leak}}$	Rate constant of $\text{Ca}^{2+}$ leak flux through the ER membrane	$1.0 \text{ s}^{-1}$
$k_{\text{pump}}$	Rate constant of ATP-ases in the ER membrane	$50.0 \mu\text{M s}^{-1}$
$k_{\text{in1}}$	Rate constant for the agonist-dependent influx into the cell	$4.0 \mu\text{M s}^{-1}$
$k_{\text{in2}}$	Constant $\text{Ca}^{2+}$ influx into the cell	$1.0 \mu\text{M s}^{-1}$
$k_{\text{out}}$	Rate constant for $\text{Ca}^{2+}$ efflux from the cell	$10.0 \text{ s}^{-1}$
$K_1$	Half-saturation constant for the $\text{IP}_3$ binding to the $\text{Ca}^{2+}$ channel	$0.2 \mu\text{M}$
$K_2$	Threshold constant for $\text{Ca}^{2+}$ pumping into the ER	$0.2 \mu\text{M}$
$K_3$	Dissociation constant of the $\text{Ca}^{2+}$ dependent component of PLC	$1.0 \mu\text{M}$
$K_4$	Activation constant of CICR	$0.69 \mu\text{M}$
$K_5$	Inhibition constant of CICR	$0.69 \mu\text{M}$
$k_+$	Maximal rate constant of $\text{IP}_3$ production	$4.0 \mu\text{M s}^{-1}$
$k_-$	Rate constant for $\text{IP}_3$ degradation by 5-phosphomonoesterase	$2.0 \text{ s}^{-1}$
$r$	Degree of cell stimulation by agonist	$0-1.0$
$a$	Amplitude of periodic forcing	$0.08-0.25 \mu\text{M s}^{-1}$

where,

$$J_{\text{ch}} = k_{\text{ch}} \left( \frac{\text{IP}_3^4}{\text{IP}_3^4 + K_1^4} \right) \times \left( \frac{K_4 \text{Ca}_{\text{cyt}}}{(\text{Ca}_{\text{cyt}} + K_4)(\text{Ca}_{\text{cyt}} + K_5)} \right)^3 \text{Ca}_{\text{er}}, \quad (4)$$

$$J_{\text{leak}} = k_{\text{leak}} \text{Ca}_{\text{er}}, \quad (5)$$

$$J_{\text{pump}} = k_{\text{pump}} \frac{\text{Ca}_{\text{cyt}}^2}{\text{Ca}_{\text{cyt}}^2 + K_2^2}, \quad (6)$$

$$J_{\text{in}} = k_{\text{in1}} \cdot r + k_{\text{in2}} \quad (7)$$

$$J_{\text{out}} = k_{\text{out}} \text{Ca}_{\text{cyt}}, \quad (8)$$

$$J_+ = k_+ \cdot r \frac{\text{Ca}_{\text{cyt}}}{\text{Ca}_{\text{cyt}} + K_3}, \quad (9)$$

$$J_- = k_- \cdot \text{IP}_3 \quad (10)$$

The model exhibits both simple and complex  $\text{Ca}^{2+}$  oscillations and hence it is suitable to be used for a comparative study of how regular and chaotic  $\text{Ca}^{2+}$  oscillations are sensitive to parameter changes and how they are flexible in response to external periodic forcing.

For the external periodic forcing a simple sinusoidal function  $f(t)$  is taken:

$$f(t) = a \sin(2\pi\nu_f t) \quad (11)$$

where  $a$  is the amplitude and  $\nu_f$  is the frequency of the external forcing. The periodic forcing is considered as a variable  $\text{Ca}^{2+}$  flux across the cell membrane. In the model this is realised by inclusion of the  $f(t)$  as additional term in Eq. (1).

All results are calculated for the parameter values given in Table 1 if not otherwise stated.

### 3. Results

The basic model system without periodic forcing expresses regular and chaotic  $\text{Ca}^{2+}$  oscillations. For better insight into the system behaviour, we calculate the largest Lyapunov exponent  $\lambda_{\text{max}}$ , which determines if the oscillatory regime is regular or chaotic. The largest Lyapunov exponent is calculated with the algorithm proposed by Wolf et al. [27]. In Fig. 1 the results are plotted vs. parameter  $r$ , which corresponds to the level of cell stimulation. The positive values of the largest Lyapunov exponent in Fig. 1 indicate that the model system behaves chaotically in two narrow parametric intervals  $0.6175 < r < 0.6205$  and  $0.6245 < r < 0.6275$ . For other parameter values in the oscillatory regime between the two subcritical Hopf bifurcations at  $r=0.2345$  (HB1 in Fig. 1) and  $r=0.6859$  (HB2 in Fig. 1), the system expresses regular periodic oscillations. Both bifur-

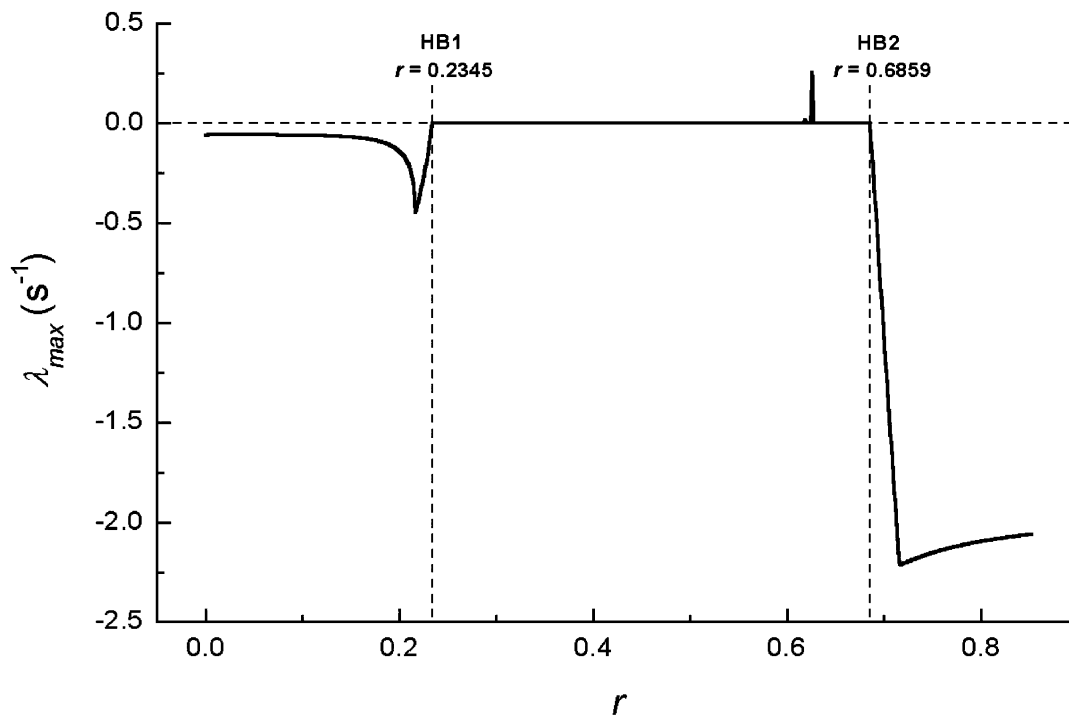


Fig. 1. The largest Lyapunov exponent  $\lambda_{\max}$  is plotted vs. parameter  $r$ .

cation points were calculated by the program XPPAUT [28].

To examine the sensitivity of  $\text{Ca}^{2+}$  oscillations in the mathematical model, we shift the parameter values given in Table 1 by  $\pm 10\%$  each at a time, and calculate the corresponding changes in model variables. To quantify changes resulting from the particular parameter shift, we take use of the response coefficients ( $R_p^X$ ), defined by the following equation (e.g. [21–23]):

$$R_p^X = \frac{\partial X/X}{\partial p/p} = \frac{p}{X} \cdot \frac{\partial X}{\partial p}, \quad (12)$$

where  $p$  is a system parameter and  $X$  is a dependent variable, like for example ionic concentrations or any of the elements of the Fourier spectrum. For better comparison of our results with that obtained previously by Galvanovskis and Sandblom [17], we take a simplified definition for the sensitivity. Since in all cases the parameters are changed for the same percent (i.e.  $\pm 10\%$ ), it holds  $\partial p/p =$

konst., and the system sensitivity ( $S_X$ ) can be simply estimated by the ratio (see also [17]):

$$S_X = \frac{X_{\text{mod}}}{X_{\text{non-mod}}}, \quad (13)$$

where  $X_{\text{mod}}$  and  $X_{\text{non-mod}}$  are values of a particular system variable after (modulated) and before (non-modulated) the parameter shift, respectively. High sensitivity corresponds to ratios  $S_X \ll 1$  and  $S_X \gg 1$ .

First, the system sensitivity regarding changes in the power of  $\text{Ca}^{2+}$  oscillations is analysed [ $X = P$  in Eq. (13)]. We shift the parameter values given in Table 1 by  $\pm 10\%$  each at a time, and calculate the corresponding ratios of the power of modulated oscillations over the power of non-modulated oscillations ( $S_P$ ). The results are presented in Table 2a. It should be noted that the power of  $\text{Ca}^{2+}$  oscillations is calculated for the oscillating signal of  $\text{Ca}_{\text{cvt}}$  with the subtracted average value of the signal. Therefore, if the ratio of the power of modulated oscillations over the

Table 2

Effects of shifting the parameter values by  $\pm 10\%$  for three different levels of cell stimulation:  $r=0.50$ ,  $r=0.62$  and  $r=0.68$ 

Modulated parameter	Regular regime at $r=0.50$		Chaotic regime at $r=0.62$		Regular regime at $r=0.68$	
	– 10%	+ 10%	– 10%	+ 10%	– 10%	+ 10%
(a)						
$k_{\text{ch}}$	0.99	1.02	1.29	0.42	1.22	0
$k_{\text{leak}}$	1.07	0.94	1.36	0.65	1.09	0.89
$k_{\text{pump}}$	0.93	1.08	0.32	1.49	0	1.43
$k_{\text{in1}}$	0.93	1.08	1.25	0.45	1.39	0
$k_{\text{in2}}$	0.96	1.05	1.25	0.51	1.17	0
$k_{\text{out}}$	1.16	0.87	0	1.19	0	1.26
$K_1$	0.93	1.05	0.36	1.52	0	1.23
$K_2$	1.17	0.83	1.73	0.02	2.44	0
$K_3$	0.95	1.05	0.47	1.44	0.72	1.16
$K_4$	0.91	1.08	0.61	1.34	0	1.35
$K_5$	0.95	1.03	0	1.45	0	1.93
$r$	0.97	0.93	1.49	0.27	4.96	0
(b)						
$k_{\text{ch}}$	0.97	1.02	1.21	5.10	0.93	0
$k_{\text{leak}}$	0.94	1.05	1.19	0.78	0.97	1.02
$k_{\text{pump}}$	1.09	0.93	5.75	1.11	0	0.90
$k_{\text{in1}}$	0.93	1.07	1.13	5.80	0.86	0
$k_{\text{in2}}$	0.97	1.03	1.21	5.03	0.97	0
$k_{\text{out}}$	1.02	0.97	0	1.21	0	0.98
$K_1$	1.12	0.89	6.11	1.08	0	0.86
$K_2$	0.90	1.10	1.02	10.1	0.67	0
$K_3$	1.12	0.91	5.70	1.10	1.11	0.90
$K_4$	0.97	1.02	4.52	1.32	0	1.04
$K_5$	1.05	0.95	0	1.1	0	0.78
$r$	0.81	1.19	0.95	6.66	0.18	0

Results are expressed (a) as ratios of the power of modulated  $\text{Ca}^{2+}$  oscillations over the power of non-modulated  $\text{Ca}^{2+}$  oscillations ( $S_p$ ), (b) as ratios of the dominant oscillation frequency of modulated  $\text{Ca}^{2+}$  oscillations over the dominant oscillation frequency of non-modulated  $\text{Ca}^{2+}$  oscillations ( $S_\nu$ ).

power of non-modulated oscillations equals zero, this means that the system has ceased oscillating altogether. In Table 2a the results are presented for parameter values  $r=0.50$ ,  $r=0.62$  and  $r=0.68$ . For parameter values  $r=0.50$  and  $r=0.62$  the results are fully in agreement with those obtained by Galvanovskis and Sandblom [17], showing that the chaotic regime at  $r=0.62$  is more sensitive than the regular one at  $r=0.50$ . On the other hand, however, the very high sensitivity of regular periodic state at  $r=0.68$  shows that in general chaotic states are not more sensitive than regular states. We further tested the sensitivity for another peri-

odic regime at  $r=0.24$ . We found that this regular periodic regime is also more sensitive than the chaotic regime at  $r=0.62$  (data not shown). Thus, our results indicate that the system sensitivity does not necessarily depend on the complexity of  $\text{Ca}^{2+}$  oscillations.

To confirm these results even further we also analyse the sensitivity of  $\text{Ca}^{2+}$  oscillations regarding changes in the oscillation frequency [ $X=\nu$  in Eq. (13)]. We shift the parameter values given in Table 1 by  $\pm 10\%$  each at a time again, and calculate the corresponding ratios of the frequency of modulated oscillations over the frequency of

non-modulated oscillations ( $S_\nu$ ). The results, showing the sensitivity of  $\text{Ca}^{2+}$  oscillations regarding changes in the oscillation frequency, are presented in Table 2b and are fully in agreement with the results presented in Table 2a. High sensitivity corresponds to values  $S_\nu \ll 1$  and  $S_\nu \gg 1$  whereas  $S_\nu = 0$  means that the system has ceased oscillating altogether. By comparing the results in Table 2a,b with Fig. 1 we see that regimes of high sensitivity coincide with the proximity of Hopf bifurcations, regardless of the complexity of  $\text{Ca}^{2+}$  oscillations.

Since our results show that chaos doesn't necessarily imply higher sensitivity of the system, it remains of interest to investigate the flexibility of chaotic and regular periodic regimes. Intuitively, chaos should be more flexible in response to external forcing since it is characterised by a variety of different amplitudes and non-harmonic frequencies. In order to examine the flexibility of  $\text{Ca}^{2+}$  oscillations in the mathematical model proposed by Shen and Larter [20] we add periodic transmembrane calcium exchange [Eq. (11)] into the model and determine the frequency range in which  $\text{Ca}^{2+}$  oscillations follow the external periodic forcing. The span of the frequency range in which  $\text{Ca}^{2+}$  oscillations are synchronised with the forcing signal represents the system ability to adapt the basic  $\text{Ca}^{2+}$  oscillations to the forcing signal. Therefore, the flexibility of the model system ( $F$ ) is quantified by the maximal span of the frequency range in which the frequency of  $\text{Ca}^{2+}$  oscillations ( $\nu$ ) equals the frequency of the external forcing ( $\nu_f$ ):

$$F = \frac{\nu_{\max} - \nu_{\min}}{\nu_0}, \quad (14)$$

where

$$\nu_{\max} = \max\{\nu, \nu = \nu_f\}, \quad (15)$$

$$\nu_{\min} = \min\{\nu, \nu = \nu_f\}, \quad (16)$$

and  $\nu_0$  is the basic oscillation frequency of the model system without periodic forcing. By varying the forcing frequency  $\nu_f$ , first  $\nu_{\max}$  and  $\nu_{\min}$  are determined by considering the condition  $\nu = \nu_f$  in Eq. (15) and Eq. (16), respectively. Then the

flexibility ( $F$ ) can be simply calculated by Eq. (14). This procedure is carried out for estimating the flexibility of different chaotic and regular oscillatory regimes. High flexibility corresponds to values  $F \gg 0$  whereas  $F = 0$  means that the system is completely inflexible.

First, the flexibility of the chaotic regime at  $r = 0.62$  is studied. The chaotic behaviour is proved by calculating the largest Lyapunov exponent, which is positive in this case (see Fig. 1). Although the regime is chaotic, a predominant frequency is well expressed as shown in Fig. 2a. The external forcing is applied to the basic  $\text{Ca}^{2+}$  oscillations. By changing the frequency of the forcing signal, we determine the range of synchronisation in which the predominant oscillation frequency of chaotic oscillations is synchronised with the forcing frequency. With high frequency forcing, we are able to enlarge the predominant frequency of the basic  $\text{Ca}^{2+}$  oscillations up to  $\nu_{\max} = 1.37 \nu_0$  (Fig. 2b). On the other hand, low forcing frequencies can reduce the basic oscillation frequency down to  $\nu_{\min} = 0.72 \nu_0$  (Fig. 2c). According to Eq. (14), this yields the flexibility of the model system  $F = 0.65$ . Below 72% and above 137% of the basic oscillation frequency, the synchronisation is lost and the frequency of  $\text{Ca}^{2+}$  oscillations tends to the value of the basic model system without forcing (see Fig. 3). Since the examined  $\text{Ca}^{2+}$  oscillations are of the bursting type, the synchronisation of the forcing signal with the main  $\text{Ca}^{2+}$  spike in Fig. 2b,c can also be seen as a quasi-phase-locking.

In the same way, we examine the flexibility of a regular periodic regime at  $r = 0.50$ . We apply the external forcing to the basic signal and determine the range of synchronisation. In contrast to the previous chaotic regime at  $r = 0.62$ , in this case we are only able to reduce the frequency of basic  $\text{Ca}^{2+}$  oscillations down to  $\nu_{\min} = 0.97 \nu_0$  and enlarge it up to  $\nu_{\max} = 1.04 \nu_0$  (see Fig. 3), which yields  $F = 0.07$ . By comparing the results obtained for the regular periodic regime at  $r = 0.50$  and for the chaotic regime at  $r = 0.62$ , one could conclude that chaotic  $\text{Ca}^{2+}$  oscillations are more flexible than regular ones. However, the question arises if this could be generalised.

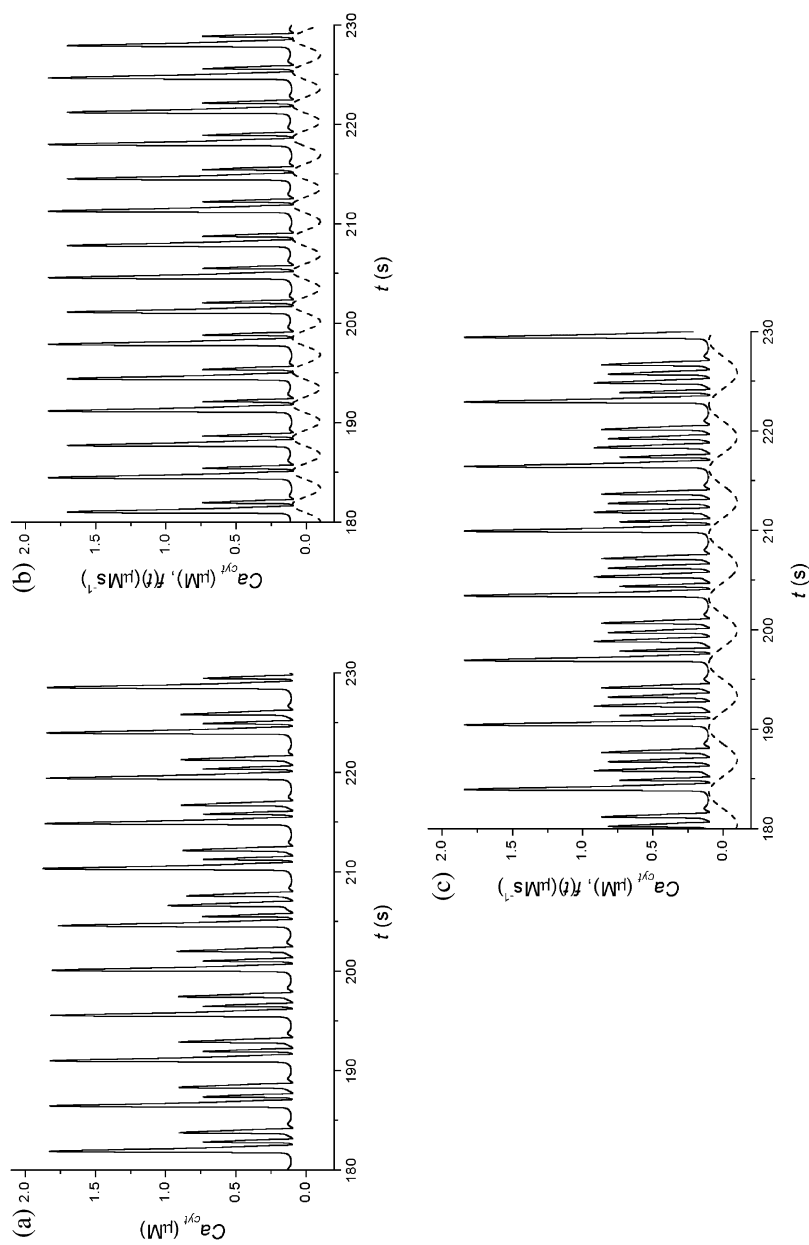


Fig. 2. Periodic forcing of chaotic  $Ca^{2+}$  oscillations at  $r=0.62$ . (a) Chaotic  $Ca^{2+}$  oscillations without periodic forcing. (b)  $Ca^{2+}$  oscillations (solid line) follow the forcing signal (dashed line) up to approximately 137% of their basic frequency. The amplitude of periodic forcing  $f(t)$  is  $a=0.1 \mu M s^{-1}$ . (c)  $Ca^{2+}$  oscillations (solid line) follow the forcing signal (dashed line) down to approximately 72% of their basic frequency. The amplitude of periodic forcing  $f(t)$  is  $a=0.1 \mu M s^{-1}$ .

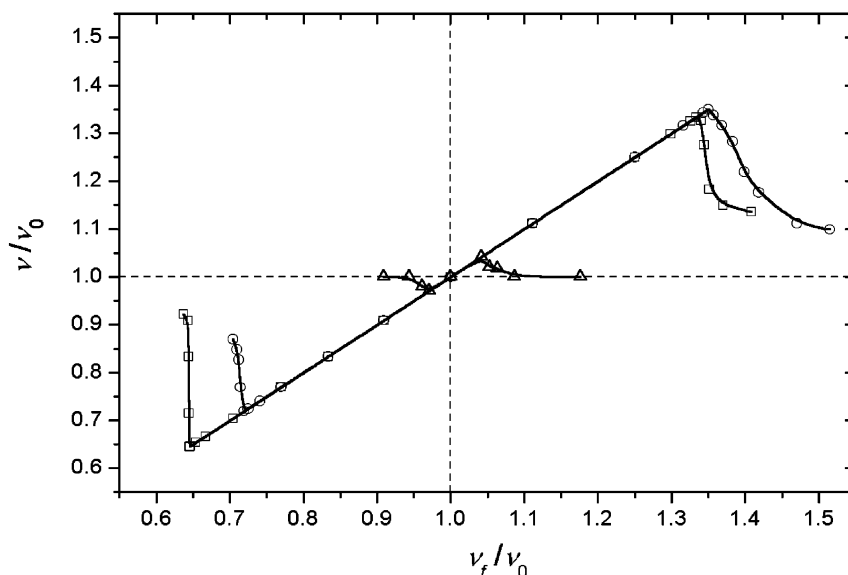


Fig. 3. Oscillation frequency ranges in which  $\text{Ca}^{2+}$  oscillations are synchronised with the forcing signal. Frequencies of forced  $\text{Ca}^{2+}$  oscillations,  $\nu$ , as well as frequencies of forcing signals,  $\nu_f$ , are normalised with respect to the frequency of basic  $\text{Ca}^{2+}$  oscillations,  $\nu_0$ . Calculations are carried out for three different values of parameter  $r$ : ( $\Delta$ )  $r=0.50$ ,  $a=0.25 \mu\text{M s}^{-1}$ , ( $\circ$ )  $r=0.62$ ,  $a=0.1 \mu\text{M s}^{-1}$  and ( $\square$ )  $r=0.624$ ,  $a=0.1 \mu\text{M s}^{-1}$ .

We investigate the flexibility of another regular  $\text{Ca}^{2+}$  oscillations at  $r=0.624$ . The span of the frequency range in which  $\text{Ca}^{2+}$  oscillations are synchronised with the forcing signal expands from  $\nu_{\min}=0.64 \nu_0$  to  $\nu_{\max}=1.33 \nu_0$  (see Fig. 3), which yields  $F=0.69$ . The result shows that this regular periodic regime is more flexible than the chaotic one at  $r=0.62$  ( $F=0.65$ ). In fact, the span of the frequency range in which  $\text{Ca}^{2+}$  oscillations are synchronised with the external forcing is larger for  $r=0.624$  than for all other parameter values tested. It should be noted that since the amplitude of  $\text{Ca}^{2+}$  oscillations slightly decreases with increasing values of parameter  $r$ , we have accordingly reduced the amplitude of forcing from  $a=0.25 \mu\text{M s}^{-1}$  (for  $r=0.50$ ) to  $a=0.1 \mu\text{M s}^{-1}$  (for  $r=0.62$  and  $r=0.624$ ). Herewith we avoid an amplitude effect of a fixed forcing signal.

Trying to explain the reasons for high flexibility of the model system, we analyse the attractive properties of the trajectories in phase space. The strength of the attraction is measured by the sum of Lyapunov exponents, which we calculate with the algorithm proposed by Wolf et al. [27]. The

sum of Lyapunov exponents corresponds to the contraction of the phase space volume and, hence to dissipation. In Fig. 4 the dissipation of the model system is plotted vs. parameter  $r$ . By comparing Fig. 4 with the results obtained in the flexibility analysis (Fig. 3), we see that regions of weak dissipation coincide with regions of high flexibility, regardless of the complexity of  $\text{Ca}^{2+}$  oscillations. Therefore, we argue that  $\text{Ca}^{2+}$  oscillations are more flexible at weak dissipation than at high dissipation.

We tested the above-suggested criterion for flexibility of  $\text{Ca}^{2+}$  oscillations on several other model systems (not shown here). We obtained qualitatively the same results, showing that the dissipation is a useful measure for the flexibility of a dynamical system. It should be noted, however, that the sum of Lyapunov exponents gives the time-averaged dissipation of the whole attractor. Therefore, for better understanding of how the system is able to deviate from the basic attractor in phase space and herewith to adapt its oscillation frequency to the oscillation frequency of the external forcing, in addition to the time-averaged dissipation of the



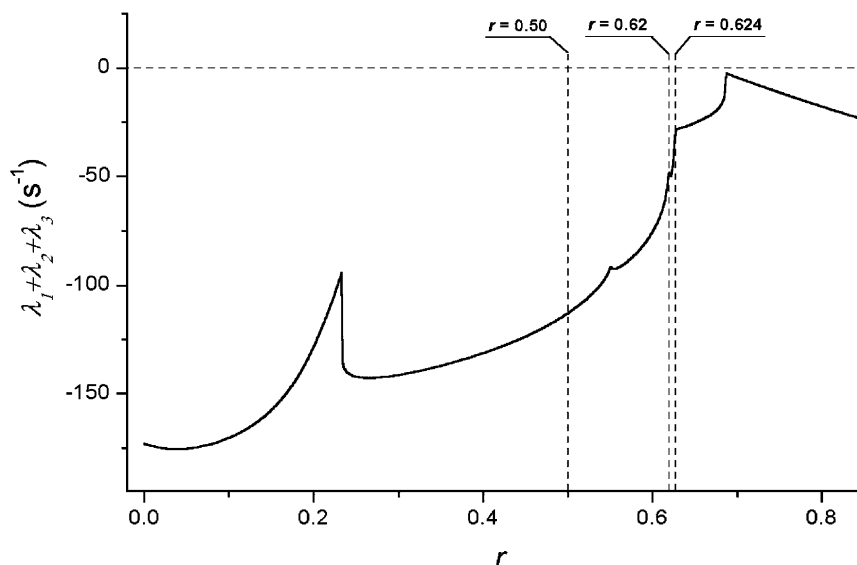


Fig. 4. Dissipation of the model system, represented as the sum of the Lyapunov exponents ( $\lambda_1 + \lambda_2 + \lambda_3$ ), is plotted vs. parameter  $r$ . Studied examples at  $r=0.50$ ,  $r=0.62$  and  $r=0.624$  are marked with vertical dashed lines.

whole attractor also the time course of local dissipation along the trajectory should be considered. We demonstrate this by examples.

Fig. 5a shows the local dissipation for periodic  $\text{Ca}^{2+}$  oscillations at  $r=0.68$ . It can be well observed that the very low time-averaged dissipation (shown in Fig. 4) results from the sinus-like time course of the local dissipation (Fig. 5a). In accordance to our previous statement, the low time-averaged dissipation implies a high flexibility of the system, which is here expressed in a high ability of the system to respond very vividly to the external periodic forcing (see Fig. 5b). The  $\text{Ca}^{2+}$  oscillations are synchronised in a large frequency range of the forcing signal, however, the amplitude of  $\text{Ca}^{2+}$  oscillations changes considerably. For biological systems, this is not fully the desired flexibility. Biological systems must not respond just vividly but also controlled and immutably, i.e. their response has to be well defined in its frequency as well as in its amplitude. To have a flexible system with well-defined responses to the external forcing, in addition to the low time-averaged dissipation, the local dissipation has to express some asymmetry. On the larger part of the

attractor, the local dissipation has to be either positive, close to zero or should oscillate around zero. This contributes to the low time-averaged dissipation and represents the flexible part of the attractor. However, in addition to this flexible part, there must also be at least one well-expressed negative dell of dissipation. This assures the system to have a strong attractive region in phase space, to which the system returns with higher probability. Herewith, the forcing signal cannot change these rigid parts of the trajectory but only alters the system behaviour in regions of low dissipation.

To demonstrate the importance of the asymmetry of local dissipation, we analyse the time course of dissipation for the highly flexible regular oscillations at  $r=0.624$ . Fig. 6a shows that in addition to the amplitude symmetric oscillations of the local dissipation around zero, two well-expressed negative dells of dissipation are present (marked with arrows in Fig. 6a). They act as stabilisers, which prevent uncontrolled vivid behaviour in response to the external forcing. The role of the attractive dells on one hand and the flexible, low attractive regions on the other hand can be demonstrated in

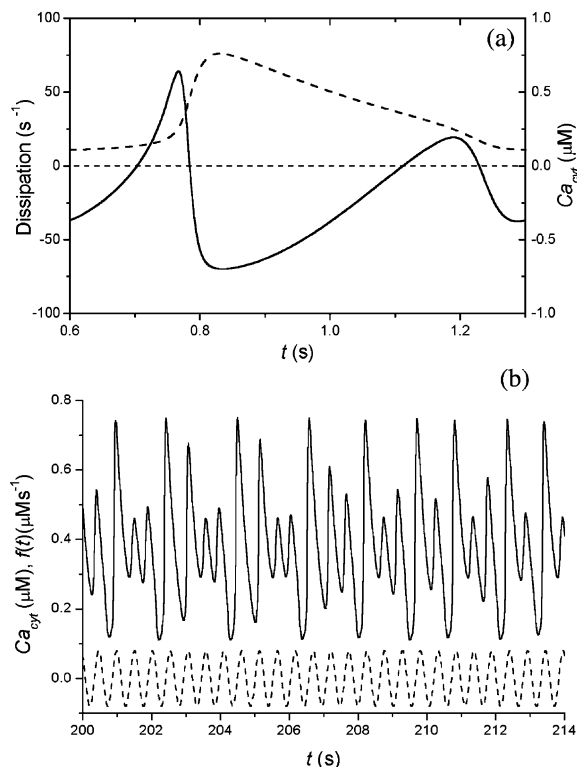


Fig. 5. Local dissipation analysis for the oscillatory regime at  $r=0.68$ . (a) Time course of the dissipation (solid line, left y-axis) and time course of  $Ca_{cyt}$  (dashed line, right y-axis) for one oscillation period. (b) Periodic forcing of regular  $Ca^{2+}$  oscillations at  $r=0.68$ . The forced  $Ca^{2+}$  oscillations (solid line) are synchronised with the forcing signal (dashed line,  $v_f=1.35v_0$ ,  $a=0.08 \mu M s^{-1}$ ); however, the amplitude of the  $Ca^{2+}$  oscillations is not well defined.

the phase space. The largest deviations of the forced trajectory from the basic attractor appear in regions of the phase space, which correspond to the most positive dissipation areas of the attractor (marked with thick lines in Fig. 6a,b). Herewith, the system is able to adjust its own frequency to the frequency of the forcing signal. However, the two well-defined negative dells (marked with arrows in Fig. 6a,b) force the system to return periodically to the strong attractive reference regions in phase space. This can be well observed in Fig. 6b where in these high attractive regions the forced trajectory coincides with the basic attractor. Taken together, we argue that a phase

space with predominantly expressed low attractive, flexible regions, and localised high attractive, rigid regions, represents a highly flexible and well-controllable system.

Our study of sensitivity and flexibility of regular and complex  $Ca^{2+}$  oscillations shows one common result, i.e. both sensitivity and flexibility do not depend on the complexity of  $Ca^{2+}$  oscillations. However, one might argue that the periodic forcing applied in studying flexibility might lead to changing from regular periodic to chaotic behaviour of

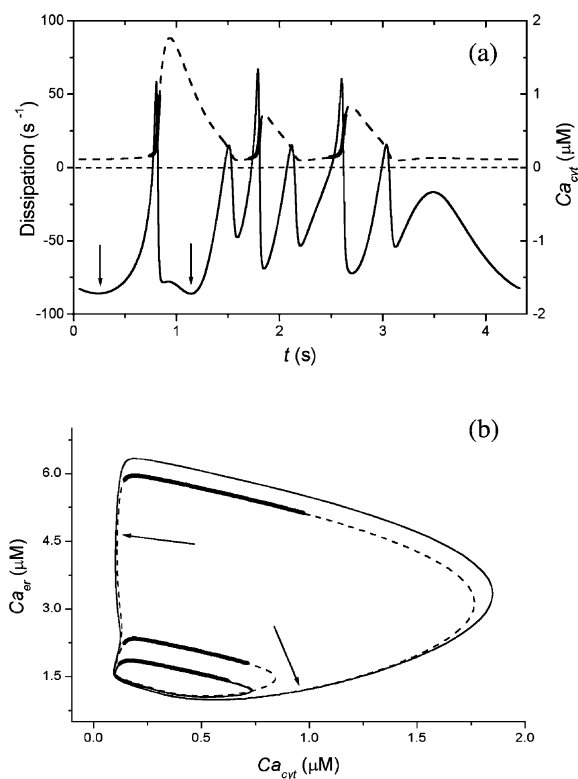


Fig. 6. Local dissipation analysis for the oscillatory regime at  $r=0.624$ . (a) Time course of the dissipation (solid line, left y-axis) and time course of  $Ca_{cyt}$  (dashed line, right y-axis) for one oscillation period. Thick line segments indicate regions of high flexibility. Arrows indicate two well-defined negative dells of local dissipation. (b) Limit cycles in 2D-phase space of  $Ca_{cyt}$  and  $Ca_{cr}$  for  $r=0.624$ . The dashed line represents the limit cycle without periodic forcing (basic attractor). The thick line segments indicate regions of high flexibility whereas arrows mark the high attractive regions with low dissipation. The solid line represents the limit cycle with external periodic forcing ( $v_f=1.25v_0$ ,  $a=0.1 \mu M s^{-1}$ ).

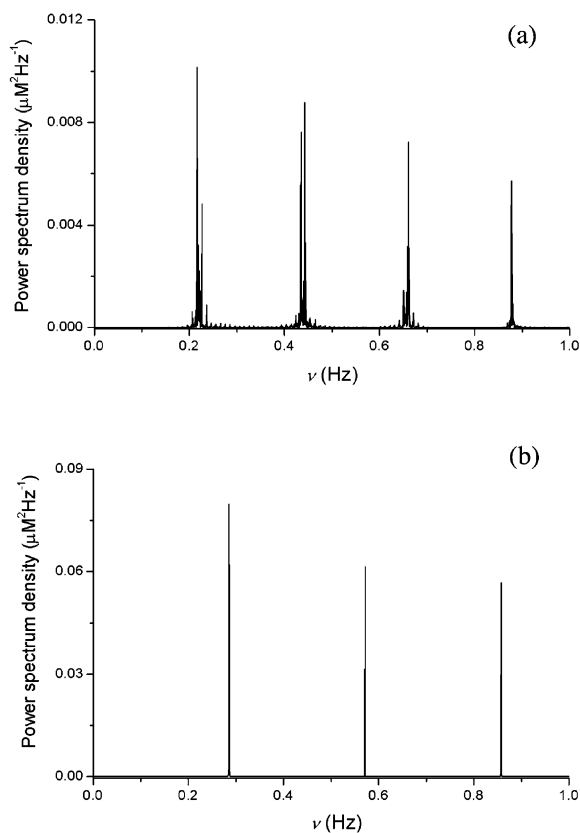


Fig. 7. Changes in power spectra caused by periodic forcing. (a) Power spectrum for chaotic  $\text{Ca}^{2+}$  oscillations at  $r=0.62$ . The spectrum corresponds to the time course of  $\text{Ca}_{\text{cyt}}$  presented in Fig. 2a. (b) Power spectrum for periodic forced  $\text{Ca}^{2+}$  oscillations. For parameter values see Table 1 and the caption of Fig. 2b.

the system. This would imply that chaos is nevertheless a necessary condition for high flexibility of the system. Therefore, in order to check regularity/irregularity of  $\text{Ca}^{2+}$  oscillations we have calculated power spectrums before and after applying the periodic forcing to the model system. The results show that in response to external periodic forcing the periodic regimes mostly remain regular whereas chaotic regimes most likely fall into a regular periodic regime. We demonstrate this by an example in Fig. 7. First, the power spectrum of unforced chaotic oscillations at  $r=0.62$  is presented in Fig. 7a. This power spectrum corresponds to the time course in Fig. 2a. Applying the periodic

forcing to these oscillations (see Fig. 2b) throws the primarily chaotic  $\text{Ca}^{2+}$  oscillations into a regular oscillatory regime. This is confirmed by the power spectrum in Fig. 7b. Similar phenomenon was also observed by Li et al. [29]. They reported about a suppression of chaotic oscillations of cyclic AMP in a suspension of *Dictyostelium discoideum* amoebae due to a small-amplitude periodic input of cyclic AMP.

#### 4. Discussion

In the paper the sensitivity and flexibility of  $\text{Ca}^{2+}$  oscillations is analysed. The study was inspired by the work of Galvanovskis and Sandblom [17] arguing that chaotic  $\text{Ca}^{2+}$  oscillations seem to be more sensitive than the periodic ones. To verify this prediction we focused our study to the same mathematical model [20] and used the same basic method for analysing the sensitivity of  $\text{Ca}^{2+}$  oscillations as proposed by Galvanovskis and Sandblom [17]. However, we examined additional examples of different periodic and chaotic regimes in order to be able to make conclusions that are more general. Furthermore, we carried out an additional sensitivity test concerning changes in frequencies of  $\text{Ca}^{2+}$  oscillations. This enabled us to give further generalisations about the sensitivity of regular periodic and chaotic  $\text{Ca}^{2+}$  oscillations. To estimate the flexibility of  $\text{Ca}^{2+}$  oscillations, we proposed a new measure. We applied the external periodic forcing to the model and measured the synchronisation range in which  $\text{Ca}^{2+}$  oscillations follow the forcing signal. By this method, we analysed the flexibility of the model system comparatively for regular and irregular chaotic  $\text{Ca}^{2+}$  oscillations. The main result of our study is that the complexity of  $\text{Ca}^{2+}$  oscillations does not directly imply higher sensitivity and flexibility of  $\text{Ca}^{2+}$  oscillations in the examined mathematical model. By several examples, we showed that regular periodic regimes could be even more sensitive and flexible than chaotic ones.

The detailed analysis shows that the sensitivity of the system strongly depends on the proximity of Hopf bifurcations, regardless of the complexity of  $\text{Ca}^{2+}$  oscillations (see Table 2a,b). However, in some cases, like in the model under consideration

[20] or in the model proposed by Borghans et al. [30], for example, complex oscillations appear in the close proximity of the Hopf bifurcations. Therefore, windows of chaotic behaviour coincide with regions of high sensitivity of the system and the results obtained earlier by Galvanovskis and Sandblom [17,18] were misleading and suggested that the higher sensitivity of the system is a consequence of the chaotical behaviour.

Intuitively, the result that the proximity of a bifurcation point determines the sensitivity of the system can be well explained. By definition, bifurcations represent large qualitative changes in the system dynamics. Therefore, if the system is near a bifurcation point, small changes in parameter values can dramatically change or even stop the oscillatory behaviour. In this case, a drastic decrease in the power as well as in the oscillation frequency appears (see Table 2a,b). From the biological point of view, this is of special importance. High sensitivity of the biological system at the threshold between a stationary state and an oscillatory regime, which is linked to a considerable change in the amplitude and/or in the frequency of  $\text{Ca}^{2+}$  oscillations, plays a crucial role assuring reliable and convincing signal transduction. The considerable changes in the amplitude and/or in the frequency of  $\text{Ca}^{2+}$  oscillations depend on the type of local or global bifurcations [31,32]. Therefore, in further more detailed sensitivity analysis the type of bifurcations should also be taken into account.

In the study of how  $\text{Ca}^{2+}$  oscillations are flexible in response to external periodic forcing, we found that the flexibility of  $\text{Ca}^{2+}$  oscillations doesn't depend on their complexity. The flexibility of  $\text{Ca}^{2+}$  oscillations in the examined model system is predominantly determined by the dissipation. Our results show that  $\text{Ca}^{2+}$  oscillations are highly flexible in regimes with weak dissipation and inflexible in regimes with high dissipation. We argue that the time-averaged dissipation for the whole attractor is a suitable index in characterizing the flexibility of the system. This result can also be well interpreted intuitively. If an attractor in form of a limit cycle is weakly attractive, it seems much easier to alter its shape, thus changing the oscillation frequency of the system.

For better understanding of the system flexibility, i.e. how much the system is able to deviate from the basic attractor in phase space and herewith to adapt its frequency to the forcing frequency, in addition to the time-averaged dissipation also the time course of local dissipation along the trajectory should be considered. We showed that a flexible system with well-defined responses to the external forcing has to express some asymmetry in the local dissipation. On larger part of the attractor, the local dissipation has to be either positive, close to zero or should oscillate around zero. This contributes to the low time-averaged dissipation and represents the flexible part of the attractor. In addition to this flexible part, there must be localised but well-expressed negative dells of dissipation. They act as stabilisers and enable well-controlled responses of  $\text{Ca}^{2+}$  oscillations to the external forcing.

Attractors that are characteristic for relaxation oscillations usually express a predominantly close to zero or sinus-like time courses of dissipation with one or more well expressed negative dells of dissipation (see e.g. Fig. 6a). Intracellular calcium oscillations with their spike-like form are in general prominent examples of relaxation oscillations [20,30,33–35]. Therefore, in most cases the flexibility of  $\text{Ca}^{2+}$  oscillations can be well determined by the time-averaged dissipation only. Our results show that the analysis of the time course of dissipation is necessary for sinus-like non-relaxation type of  $\text{Ca}^{2+}$  oscillations. Although this type of  $\text{Ca}^{2+}$  oscillations is of moderate physiological importance, it appears in the majority of mathematical models. However, in the models the sinus-like  $\text{Ca}^{2+}$  oscillations are mainly a consequence of soft excitation [32,36]. Therefore, they are restricted to small parameter ranges close to special types of bifurcations and herewith have only very limited biological importance.

Our results indicate that dissipation should be considered in analysing the sensitivity and flexibility of  $\text{Ca}^{2+}$  oscillations. Also from the biological point of view, the relation between low dissipation and high flexibility of  $\text{Ca}^{2+}$  oscillations seems to be reasonable, since in view of low free energy consumption, dissipation of biological systems should be minimised [37]. However, it should

be noted that our study was made for a rather simple mathematical model [20]. Although we have additionally tested the sensitivity and flexibility criteria on some more complex mathematical models for  $\text{Ca}^{2+}$  oscillations [30,34,35] and obtained qualitatively the same results (not shown in the paper), further studies will be necessary in order to give more general conclusions. It would be interesting to find mathematical models in which the sensitivity and flexibility criteria considerably deviate from that suggested in this paper. This would give new insights into the system properties that determine high sensitivity and flexibility of dynamical systems. In further studies, it seems promising to analyse not only the attractive properties of a given limit cycle but also the attractive properties of the whole surrounding phase space. In this way, we would get a topological picture of the landscape in which the limit cycle lies, which would drastically improve our understanding of the sensitivity, flexibility and robustness of  $\text{Ca}^{2+}$  oscillations.

## References

- [1] M.J. Berridge, M.D. Bootman, P. Lipp, Calcium—a life and death signal, *Nature* 395 (1998) 645–648.
- [2] G. Dupont, S. Swillens, C. Clair, T. Tordjmann, L. Combettes, Hierarchical organisation of calcium signals in hepatocytes: from experiments to models, *Biochim. Biophys. Acta* 1498 (2000) 134–152.
- [3] S. Schuster, M. Marhl, T. Höfer, Modelling of simple and complex calcium oscillations. From single-cell responses to intercellular signalling, *Eur. J. Biochem.* 269 (2002) 1333–1355.
- [4] W.-h. Li, J. Llopis, M. Whitney, G. Zlokarnik, R.Y. Tsien, Cell-permeant caged  $\text{InsP}_3$  ester shows that  $\text{Ca}^{2+}$  spike frequency can optimise gene expression, *Nature* 392 (1998) 936–941.
- [5] P. De Koninck, H. Schulman, Sensitivity of CaM kinase II to the frequency of  $\text{Ca}^{2+}$  oscillations, *Science* 279 (1998) 227–230.
- [6] R.E. Dolmetsch, K. Xu, R.S. Lewis, Calcium oscillations increase the efficiency and specificity of gene expression, *Nature* 392 (1998) 933–936.
- [7] M. Marhl, S. Schuster, M. Brumen, Mitochondria as an important factor in the maintenance of constant amplitudes of cytosolic calcium oscillations, *Biophys. Chem.* 71 (1998) 125–132.
- [8] V. Grubelnik, A.Z. Larsen, U. Kummer, L.F. Olsen, M. Marhl, Mitochondria regulate the amplitude of simple and complex calcium oscillations, *Biophys. Chem.* 94 (2001) 59–74.
- [9] K.U. Bayer, P. De Koninck, H. Schulman, Alternative splicing modulates the frequency-dependent response of CaMKII to  $\text{Ca}^{2+}$  oscillations, *EMBO J.* 21 (2002) 3590–3597.
- [10] A. Hudmon, H. Schulman, Structure-function of the multifunctional  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase II, *Biochem. J.* 364 (2002) 593–611.
- [11] K. Prank, F. Gabbiani, G. Brabant, Coding efficiency and information rates in transmembrane signalling, *Biosystems* 55 (2000) 15–22.
- [12] K. Prank, M. Kropp, G. Brabant, Humoral coding and decoding Complexity Biology, Inform, Processing Novartis Foundation Symposium 239 (2001) 96–110.
- [13] C. Suguna, K.K. Chowdhury, S. Sinha, Minimal model for complex dynamics in cellular processes, *Phys. Rev. Part E* 60 (1999) 5943–5949.
- [14] U. Kummer, G. Baier, L.F. Olsen, Robustness in a model for calcium signal transduction dynamics, in: J.H.S. Hofmeyr, J.M. Rohwer, J.L. Snoep (Eds.), *Animating the Cellular Map*, Stellenbosch University Press, Stellenbosch, 2000, pp. 171–176.
- [15] L. Läger, M. Kloppstech, C. Schöfl, T.J. Sejnowski, G. Brabant, K. Prank, Noise enhanced hormonal signal transduction through intracellular calcium oscillations, *Biophys. Chem.* 91 (2001) 157–166.
- [16] S. Zhong, F. Qi, H. Xin, Internal stochastic resonance in a model for intracellular calcium oscillations, *Chem. Phys. Lett.* 342 (2001) 583–586.
- [17] J. Galvanovskis, J. Sandblom, Periodic forcing of intracellular calcium oscillators. Theoretical studies of the effects of low frequency fields on the magnitude of oscillations, *Bioelectrochem. Bioenerg.* 46 (1998) 161–174.
- [18] J. Sandblom, J. Galvanovskis, Electromagnetic field absorption in stochastic cellular systems: enhanced signal detection in ion channels and calcium oscillators, *Chaos Solitons Fractals* 11 (2000) 1905–1911.
- [19] A. Goldbeter, G. Dupont, M.J. Berridge, Minimal model for signal-induced  $\text{Ca}^{2+}$  oscillations and for their frequency encoding through protein phosphorylation, *Proc. Natl. Acad. Sci. USA* 87 (1990) 1461–1465.
- [20] P. Shen, R. Larter, Chaos in intracellular  $\text{Ca}^{2+}$  oscillations in a new model for non-excitable cells, *Cell Calcium* 17 (1995) 225–232.
- [21] R. Heinrich, S. Schuster, *The regulation of cellular systems*, Chapman & Hall, New York, 1996.
- [22] B.N. Kholodenko, O.V. Demin, H.V. Westerhoff, Control analysis of periodic phenomena in biological systems, *J. Phys. Chem. Part B* 101 (1997) 2070–2081.
- [23] K.A. Reijenga, H.V. Westerhoff, B.N. Kholodenko, J.L. Snoep, Control analysis for autonomously oscillating biochemical networks, *Biophys. J.* 82 (2002) 99–108.
- [24] R. Larter, B. Aguda, C. Bush, T. Lonis, Multiple steady states, complex oscillations, and the devil's staircase in the peroxidase-oxidase reaction, *J. Chem. Phys.* 87 (1987) 5765–5771.

- [25] T. Meyer, L. Stryer, Molecular model for receptor-stimulated calcium spiking, *Proc. Natl. Acad. Sci. USA* 85 (1988) 5051–5055.
- [26] L. Stryer, T. Meyer, Calcium spiking, *Annu. Rev. Biophys. Biophys. Chem.* 20 (1991) 153–174.
- [27] A. Wolf, J.B. Swift, H.L. Swinney, J.A. Vastano, Determining Lyapunov exponents from a time series, *Physica D* 16 (1985) 285–317.
- [28] G.B. Ermentrout, XPPAUT: The differential equation solving tool, <http://www.math.pitt.edu/~bard/xpp/xpp.html> (1996).
- [29] Y. Li, J. Halloy, J. Martiel, B. Wurster, A. Goldbeter, Suppression of chaos by periodic oscillations in a model for cyclic-AMP signalling in dictyostelium cells, *Experientia* 48 (1992) 603–606.
- [30] J.A.M. Borghans, G. Dupont, A. Goldbeter, Complex intracellular calcium oscillations. A theoretical exploration of possible mechanisms, *Biophys. Chem.* 66 (1997) 25–41.
- [31] S.H. Strogatz, *Non-linear Dynamics and Chaos With Applications to Physics, Biology, Chemistry, and Engineering*, Perseus Publishing, Cambridge, 1994.
- [32] S. Schuster, M. Marhl, Bifurcation analysis of calcium oscillations: Time-scale separation, canards and frequency lowering, *J. Biol. Syst.* 9 (2001) 291–314.
- [33] G. Houart, G. Dupont, A. Goldbeter, Bursting, chaos and birhythmicity originating from self-modulation of the Inositol 1,4,5-trisphosphate signal in a model for intracellular  $\text{Ca}^{2+}$  oscillations', *Bull. Math. Biol.* 61 (1999) 507–530.
- [34] M. Marhl, T. Haberichter, M. Brumen, R. Heinrich, Complex calcium oscillations and the role of mitochondria and cytosolic proteins, *BioSystems* 57 (2000) 75–86.
- [35] U. Kummer, L.F. Olsen, C.J. Dixon, A.K. Green, E. Bornberg-Bauer, G. Baier, Switching from simple to complex oscillations in calcium signalling, *Biophys. J.* 79 (2000) 1188–1195.
- [36] A. Goldbeter, *Biochemical oscillations and cellular rhythms*, Cambridge University Press, Cambridge, 1996.
- [37] J.W. Stucki, M. Compiani, S.R. Caplan, Efficiency of energy conversion in model biological pumps. Optimisation by linear non-equilibrium thermodynamics relations, *Biophys. Chem.* 18 (1983) 101–109.